Preparation for the exam 2

Chem112L, Spring 2012

Exam date: Monday, June 11, 4 PM

This exam focuses on mass spectrometry, protein-ligand interactions as studied by spectrophotometry, and crystallization / diffraction analysis of macromolecules. I expect that you know the basic material from the three previous experiments as well. I intend to have a mix of knowledge-showing essay-type, problem-solving, and multiple-choice questions. Knowledge of the following helps you in preparing for the exam:

- 1. Physical principles behind each of the molecular process
 - a. Protein ionization and its relevance to mass spectrometry
 - b. Ligand binding to macromolecules
 - c. Protein crystallization
- 2. Physical principles behind each of the observation/detection methods
 - a. X-ray diffractiometry
 - b. UV absorption spectroscopy, design of a UV-Vis spectrophotometer
 - c. Separation of molecules based on m/z values; protein identification
 - d. Identification of structure of peptides and metabolites via mass spectrometry
- 3. Theoretical description of biochemical processes such as ligand binding
 - a. Estimation of thermodynamic parameters from experimental data
 - b. Dissociation constant in relation to equilibrium concentrations
 - c. Rate constants in relation to the equilibrium constant
 - c. Relationships between free energy, enthalpy, entropy and heat capacity
- 4. Structural and functions concepts pertaining to ligand binding and protein function
 - a. Biological function and catalytic mechanism of lysozyme
 - b. Structure of peptides, proteins, and nucleic acids
 - c. Electronic structure of molecules, molecular orbitals
 - d. Electronic $\pi \rightarrow \pi^*$, and $n \rightarrow \pi^*$ transitions
 - e. Solvatochromic shifts in UV-Vis spectra
- 5. Instrumentation
 - a. Basic design and operation of a X-ray diffractiometers and synchrotrons
 - b. Basic design and operation of a dual-beam UV spectrophotometer
 - c. Ionization/vaporization methods for small molecule MS
 - d. Ionization/vaporization methods for macromolecule MS
 - e. Basic design and operation of a ESI mass spectrometer
 - f. Comparison of different mass analyzers
 - g. Operation of a SLR digital camera for image acquisition
- 6. Broader applications of methods covered; other approaches to study these phenomena
 - a. Using mass spectrometry to study ligand binding
 - b. Other UV spectrophotometry to study structure and ionization of binding
 - c. Comparison of NMR and crystallography for determination of protein structures
 - d. How to apply crystallography to other problems in biochemistry
 - e. Applications of mass spectrometry in proteomics

7. Practical aspects of each of the experiments and computations

- a. Why such wavelengths and cuvettes
- b. Why such concentrations, pH, salts, buffers, etc
- 8. Data analysis.
 - a. Understand why we used such model equations for fitting
 - b. Understand the meaning of each of the fitting parameters
 - c. Understand the measures of quality of data and fitting
 - d. Understand the workflow of scientific data analysis programs such as Mathematica
 - e. Understanding the workflow of a crystallography programs such as Coot
 - f. Understand how to interpret 2D X-ray diffraction data
 - g. Understand how to interpret 3D electron density maps
 - h. Different ways to plot and analyze ligand binding data
 - i. Different ways to plot ligand binding data
 - j. Analysis of binding data when ligand and protein concentrations are similar
 - k. Analysis of first order irreversible kinetics
 - 1. Analysis of first order consecutive reaction kinetics

9. Miscellaneous

- a. How to derive equations for association/dissociation equilibrium
- b. Fluorescence as a method to study ligand binding
- c. Assumption of high ligand and low protein concentration
- d. How to visually estimate fitting parameters
- e. How to derive equations that describe temperature-dependence of equilibrium constants

Answers to many of the questions require substantial thinking. Memorizing all the material may not be the best way to study for this exam.